



Diagnostics for Life

Avioq HIV-1 Technical Tips

1. Bring all reagents to room temperature (15 - 30°C) before use.
2. The ABTS Substrate Diluent should be at room temperature before reconstituting the ABTS. This will minimize auto-oxidation of the ABTS.
3. Check incubator temperature before use. Incubator temperature should be stable at $37 \pm 2^{\circ}\text{C}$.
4. Make sure that all strips are secure in the holder before reagent addition.
5. Hold the multichannel pipet at a 75° angle to the strip wells. Do not touch the bottom of the well with the pipet tip.
6. After manual pipetting of sample and reagents, carefully **BLOT** the surface of the microelisa plate with an absorbent paper to remove moisture from the tray and well rims.
7. **Do not** reuse reagent troughs.
8. The wash procedure is critical. Incomplete washing will adversely affect the test outcome. Make sure that the washer is filling and aspirating properly before use (see package insert "wash procedure"). Routine preventive maintenance of the wash system is strongly recommended.
9. Firmly **TAP** the microelisa plate on absorbent paper after the last aspiration of the wash cycle if excess wash solution is present.
10. Do not allow the microelisa test wells to dry once the assay is started. Fill the wells with the next required reagent immediately (within 10 minutes) after washing.
11. Before reading the microelisa plate, wipe the bottom of the plate to remove any excess wash or glove powder.
12. Carefully place the plate on the reader carrier. Try not to "snap" the plate into the carrier; this could cause splashing.
13. Visually check that the wells that are interpreted as reactive are green. If discrepancies are noted, check the bottom of the wells for liquid, glove powder, or other material.
14. Use only Hamilton brand tips when using the Hamilton Microlab 530b Pipetter/Diluter.