



Diagnostics for Life

Avioq HTLV-I/II Technical Tips

1. Ensure all reagents to room temperature (15-30°C) before use. The reconstituted EnzAbody Concentrate should be brought to room temperature 5-10 minutes prior to working conjugate preparation and returned to 2-8°C after use.
2. The TMB Substrate and the EnzAbody Working Solution should be prepared in disposable polypropylene containers. Do not use polystyrene containers.
3. Upon opening a new kit, reconstitute the EnzAbody Concentrate first. Allow at least 30 minutes at room temperature for the lyophilized cake to dissolve. Invert to mix, examine for complete dissolution. For optimal dissolution of conjugate prepare the day before testing.
4. Ensure that any crystals and/or precipitate in the Wash Buffer Concentrate are completely dissolved. Use a warm water bath or 37°C incubator to accelerate this process. Use only Wash Buffer Concentrate formulated for Avioq HTLV-I/II.
5. Check incubator before use. Incubator temperature should be stable at $37 \pm 2^\circ\text{C}$.
6. Make sure that all strips are secure in the holder before reagent addition.
7. Hold the multichannel pipet at a 75° angle to the strip wells. Do not touch the bottom of the well with the pipet tip.
8. Mix test specimens completely. The Positive and Negative Controls may be vortexed before pipetting.
9. After manual pipetting of sample and reagents, carefully **BLOT** the surface of the microelisa plate with an absorbent paper to remove moisture from the tray and well rims before applying the plate sealer.
10. Change gloves prior to handling EnzAbody Concentrate or Working EnzAbody Conjugate. When preparing the Working EnzAbody Conjugate: Use a calibrated pipette. Refer to manufacturer's instructions for optimal pipetting of "TD" or "TC" pipettes. Mix conjugate several times prior to aspirating desired volume. Dispense measured volume of conjugate concentrate into polypropylene or clean glass vial containing conjugate diluent. Invert several times to completely mix.
11. **Do not** reuse reagent troughs.
12. Make up only enough TMB Substrate solution for immediate use. Do not handle TMB Substrate or reagent trough with gloves that have come into contact with bleach or EnzAbody.
13. Firmly **TAP** the microelisa plate on absorbent paper after the last aspiration of the wash cycle if excess wash solution is present.
14. Do not allow the microelisa test wells to dry once the assay is started. Fill the wells with the next required reagent immediately (within 10 minutes) after washing.
15. Before reading the microelisa plate, wipe the bottom of the plate to remove any excess wash or glove powder.
16. Carefully place the plate on the reader carrier. Try not to "snap" the plate into the carrier; this could cause splashing.
17. Visually check that the wells that are interpreted as reactive are yellow. If discrepancies are

noted, check the bottom of the wells for liquid, glove powder, or other material.

18. Use only Hamilton brand tips when using the Hamilton Microlab 530b Pipetter/Diluter.

Preparing Manual in-Well Dilutions (Direct Sample Addition)

1. Pipettes calibrated to deliver the required volumes must be used.
2. When sample is added to the well it should be thoroughly mixed with Sample Diluent (at least 5 times).
3. When mixing the sample with the Sample Diluent, it is important to avoid the creation of bubbles in the well.
4. After mixing has been completed be sure to remove the pipette tip from the well before releasing the plunger.
5. Mix all samples well prior to pipetting.
6. Within 30 minutes of sample/control addition, incubate at $37 \pm 2^{\circ}\text{C}$ for 60 ± 5 minutes.
7. Ensure that the amount of sample drawn into the pipette is correct.

Note: refer to package insert for Indirect Sample Addition

Conjugate Storage and Preparation

1. Change gloves prior to opening vial of EnzAbody Concentrate.
2. Keep EnzAbody Concentrate at $2-8^{\circ}\text{C}$ until just prior to preparing working solution. Immediately return conjugate concentrate to $2-8^{\circ}\text{C}$.
3. After reconstitution mix thoroughly by inversion. Allow to stand 30 minutes at room temperature to dissolve. Store at $2-8^{\circ}\text{C}$.
4. To ensure the stopper does not become contaminated with serum, clean the bench top and change gloves prior to handling EnzAbody.
5. Prepare EnzAbody Working Solution in polypropylene vessels. Do not use polystyrene. The EnzAbody Working Solution is stable for 4 hours at room temperature.